

Tree species delimitation in tropical forest inventories: perspectives from a taxonomically challenging case study

Paulo Henrique Gaem^{a,*}, Ana Andrade^b, Fiorella Fernanda Mazine^a & Alberto Vicentini^c

^a Grupo de Pesquisa em Ecologia, Sistemática e Conservação de Recursos Naturais, Universidade Federal de São Carlos, Campus Sorocaba, Rodovia João Leme dos Santos (SP-264), km 110, 18052-780, Sorocaba, SP, Brazil

^b Projeto Dinâmica Biológica de Fragmentos Florestais, Instituto Nacional de Pesquisas da Amazônia, Avenida André Araújo, 2936, 69067-375, Manaus, AM, Brazil

^c Laboratório de Botânica Amazônica, Programa de Pós-Graduação em Botânica, Instituto Nacional de Pesquisas da Amazônia, Avenida André Araújo, 2936, 69067-375, Manaus, AM, Brazil

* Corresponding author

Supplementary File 3. Programming routine used to data preparation and statistical analyses in R environment.

```
#Tested classifications
```

```
classificacoes = c("class_paulo", "class_aleatoria")
```

```
#Dataset
```

```
file=read.table('file.csv', sep='/t', as.is=T, header=T);
```

```
rownames(file)=file$id_1
```

1. Stepwise procedure with spectral data

```
#Selection of data used in stepwise procedure
```

```
identifier=file$id_2, names=file$morph_class, nirdad=file[,16:1572]
```

```
espectros=cbind(identifier, names, nirdad)
```

```
#Remove species with one sample only
```

```
library(dplyr)
```

```
especs_once=count(espectros, espectros$names)
```

```

especs_once=subset(especs_once, especs_once$n==1)
especs_more=espectros[!espectros$names %in% especs_once$`espectros$names`,]
detach("package:dplyr", unload=TRUE)
#Stepwise procedure itself
library(MASS)
library(klaR)
x=as.matrix(nirdad); grouping=names; method='lda'; direction='both'; criterion='CR'; fold=10;
maxvar=floor(nrow(x)/3);
stepwise.res = stepclass(x, grouping, method, maxvar = maxvar, direction = direction, criterion =
criterion, fold = fold, output = F, min1var = TRUE)
formula=sc_obj$formula
vars=all.vars(formula)
vars=vars[-1]
nirsel=nirdad[, (names(nirdad) %in% formula)]

```

2. PCA-reduction of spectral data

```

nirpca = prcomp(nirdad)
nirpca = nirpca$x[,1:129]
nirpca = as.data.frame(nirpca,stringsAsFactors = F)
rownames(nirpca) = rownames(nirdad)

```

3. Morphometric data

```

morph=file[,5:15]

```

4. Spectral plus morphometric datasets

```

rn = rownames(morph)
nir_morph = cbind(nirdad[rn,],morph)
nirsel_morph = cbind(nirsel[rn,],morph)
rn = rownames(morph)

```

```
nirpcamorph = cbind(nirpca[rn,],morph)
```

5. Define a leave-one-out cross-validation LDA

```
library("caret")
```

```
myControl_LOOCV <- trainControl(
```

```
  method = "LOOCV",
```

```
  ## outras definicoes
```

```
  # imprime iteracao na tela?
```

```
  verboseIter = TRUE,
```

```
  # salva as predicoes
```

```
  savePredictions = TRUE,
```

```
  # salva a probabilidade das classes
```

```
  classProb = TRUE
```

```
)
```

6. Calculate the models' accuracies for the morphotype-based classification and the aleatory classification

```
resultado.logcv = NULL
```

```
cl=1
```

```
for(cl in 1:length(classificacoes)) {
```

```
  set.seed(453)
```

```
  classe = classificacoes[cl]
```

```
  ln = rownames(nirdad)
```

```
  grouping = as.vector(nomes_morph[ln,classe])
```

```
  tbl = table(grouping)
```

```
  tira = which(grouping%in%names(tbl[tbl<3]))
```

```
print(paste(classe,"teste NIR tirado as especies:",paste(grouping[tira],collapse = " "))  
lda1 <- train(x=nirdad[-tira,],y=grouping[-tira],method="lda",trControl = myControl_LOOCV)  
n2 = paste(classe,"nirdad",sep= ".")  
print(paste(n2,"ACCURACY: ",round(lda1$results$Accuracy,2)*100,"% ",sep=""))  
resultado.logcv[[n2]] = lda1
```

```
ln = rownames(morph)  
grouping = as.vector(nomes_morph[ln,classe])  
tbl = table(grouping)  
tira = which(grouping%in%names(tbl[tbl<3]))  
print(paste(classe,"teste morph tirado as especies:",paste(grouping[tira],collapse = " "))  
lda2 <- train(x=morph[-tira,],y=grouping[-tira],method="lda",trControl = myControl_LOOCV)  
n2 = paste(classe,"morph",sep= ".")  
print(paste(n2,"ACCURACY: ",round(lda2$results$Accuracy,2)*100,"% ",sep=""))  
resultado.logcv[[n2]] = lda2
```

```
ln = rownames(nir_morph)  
grouping = as.vector(nomes_morph[ln,classe])  
tbl = table(grouping)  
tira = which(grouping%in%names(tbl[tbl<3]))  
print(paste(classe,"teste nir+morph tirado as especies:",paste(grouping[tira],collapse = " "))  
lda3 <- train(x=nir_morph[-tira,],y=grouping[-tira],method="lda",trControl =  
myControl_LOOCV)  
n2 = paste(classe,"nir+morph",sep= ".")  
print(paste(n2,"ACCURACY: ",round(lda3$results$Accuracy,2)*100,"% ",sep=""))  
resultado.logcv[[n2]] = lda3
```

```
ln = rownames(nirsel)
grouping = as.vector(nomes_morph[ln,classe])
tbl = table(grouping)
tira = which(grouping%in%names(tbl[tbl<3]))
print(paste(classe,"teste NIRsel tirado as especies:",paste(grouping[tira],collapse = " "))
lda4 <- train(x=nirsel[-tira,],y=grouping[-tira],method="lda",trControl = myControl_LOOCV)
n2 = paste(classe,"nirsel",sep= ".")
print(paste(n2,"ACCURACY: ",round(lda4$results$Accuracy,2)*100,"%",sep=""))
resultado.logcv[[n2]] = lda4
```

```
ln = rownames(nirsel_morph)
grouping = as.vector(nomes_morph[ln,classe])
tbl = table(grouping)
tira = which(grouping%in%names(tbl[tbl<3]))
print(paste(classe,"teste nirsel+morph tirado as especies:",paste(grouping[tira],collapse = " "))
lda5 <- train(x=nirsel_morph[-tira,],y=grouping[-tira],method="lda",trControl =
myControl_LOOCV)
n2 = paste(classe,"nirsel+morph",sep= ".")
print(paste(n2,"ACCURACY: ",round(lda5$results$Accuracy,2)*100,"%",sep=""))
resultado.logcv[[n2]] = lda5
```

```
ln = rownames(nirpca)
grouping = as.vector(nomes_morph[ln,classe])
tbl = table(grouping)
tira = which(grouping%in%names(tbl[tbl<3]))
```

```

print(paste(classe,"teste nirPCA tirado as especies:",paste(grouping[tira],collapse = " "))
lda6 <- train(x=nirpca[-tira,],y=grouping[-tira],method="lda",trControl = myControl_LOOCV)
n2 = paste(classe,"nirPCA",sep= ".")
print(paste(n2,"ACCURACY: ",round(lda6$results$Accuracy,2)*100,"%",sep=""))
resultado.logcv[[n2]] = lda6

ln = rownames(nirpcamorph)
grouping = as.vector(nomes_morph[ln,classe])
tbl = table(grouping)
tira = which(grouping%in%names(tbl[tbl<3]))
print(paste(classe,"teste nirPCA+morph tirado as especies:",paste(grouping[tira],collapse = " "))
lda7 <- train(x=nirpcamorph[-tira,],y=grouping[-tira],method="lda",trControl =
myControl_LOOCV)
n2 = paste(classe,"nirPCA+morph",sep= ".")
print(paste(n2,"ACCURACY: ",round(lda7$results$Accuracy,2)*100,"%",sep=""))
resultado.logcv[[n2]] = lda7
}

```

7. Save Rdata

```
save(resultado.logcv,file="testemorfotipo.Rdata")
```

8. Calculate accuracies

```

load("testemorfotipo_indet16.Rdata")
pegaaccuracia <- function(x) {
  round(x$results$Accuracy,2)
}
accuracias <- sapply(resultado.logcv,pegaaccuracia)
names(accuracias)=names(resultado.logcv)

```

```

acuracias = as.data.frame(acuracias,stringsAsFactors=F)

colnames(acuracias) = "Acuracia"

acuracias

write.table(acuracias,sep="\t",na="",quote=T,row.names = T)

```

9. Confusion matrixes

```

load("testemorfotipo.Rdata")

abc=resultado.logcv$'classification'$pred[,1:2]

plotmatriz <- function(matriz,bg.cols,txt.cols, valcex=1, cexaxis=1) {

  tb = matriz

  xx = 1:ncol(tb)

  yy = seq(1,nrow(tb),length.out=length(xx))

  plot(xx,yy,type='n',xaxt='n',yaxt='n',xlab="",ylab="", pty='m',

xlim=c(0.5,max(xx)+.5),ylim=c(0.5,max(yy)+0.5))

  ay = 1:ncol(tb)

  ax = 1:nrow(tb)

  abline(v=ay,h=ax,lty='dotted',lwd=0.5)

  nn = rownames(tb)

  nncl = colnames(tb)

  for(l in 1:nrow(tb)) {

    for(cc in 1:ncol(tb)) {

      cl = bg.cols[l,cc]

      txtc = txt.cols[l,cc]

      val = tb[l,cc]

      if (!is.na(val)) {

        rect(xleft=cc-0.5,ybottom=l-0.5,xright=cc+0.5,ytop=l+0.5,density=-1,border=NA,col=cl)

        text(cc,l,labels=val,cex=valcex,col=txtc)

```

```

    }
  }
}
axis(side=3,at=ay,labels=colnames(tb),cex.axis=cexaxis,las=2)
axis(side=2,at=ax,labels=rownames(tb),cex.axis=cexaxis,las=2)
}
tb = table(abc$obs,abc$pred)
tb = as.matrix(tb)
cl = rownames(tb)
cl2 = colnames(tb)
falta = cl[!cl%in%cl2]
if (length(falta)>0) {
  mm = matrix(0,nrow=nrow(tb),ncol=length(falta),dimnames = list(rownames(tb),falta))
  tb = cbind(tb,mm)
  rn = sort(rownames(tb),decreasing = F)
  rn2 = sort(rownames(tb),decreasing = F)
  tb = tb[rn,rn2]
}
bg.cols= tb
txt.cols = tb
unique(as.vector(tb))
bg.cols[tb==0] = gray(level=1)
txt.cols[tb==0] = gray(level=1)
bg.cols[tb>0 & tb<=2] = gray(level=0)
txt.cols[tb>0 & tb<=2] = gray(level=1)
bg.cols[tb>2] = gray(level=0)

```

```
txt.cols[tb>2] = gray(level=1)

tb[tb==0] = NA

fn = paste("confusion_matrix.png")

png(file=fn, width = 21, height = 21, units = "cm", pointsize = 10,bg = "white", res = 600)

par(mar=c(2,10,10,2))

plotmatriz(tb,bg.cols=bg.cols,txt.cols=txt.cols,valcex=0.8, cexaxis = 0.8)

dev.off()
```